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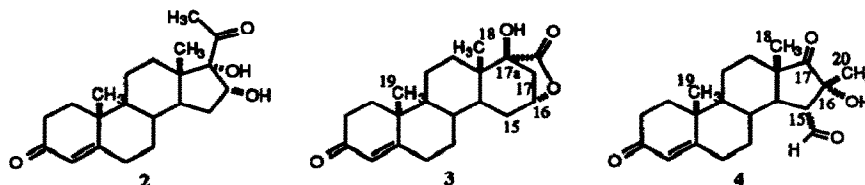
## Novel Steroids from Cetyltrimethylammonium Permanganate-Initiated Oxidative Rearrangements of 16-Dehydroprogesterone

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**Abstract:** Cetyltrimethylammonium permanganate initiates oxidative opening of the D-ring of 16-dehydroprogesterone in  $\text{CH}_2\text{Cl}_2$ . The novel steroids 3 and 4 have been isolated and characterized as the major products obtained from oxidative rearrangement of the steroid backbone. The X-ray crystal structure of the D-homosteroid 3 is provided, and a putative mechanism that invokes a benzilic acid rearrangement for formation of 3 is presented.

In the course of our development of photoaffinity labeling reagents for the progesterone receptor, we required  $16\alpha,17\alpha$ -dihydroxyprogesterone (2).<sup>1</sup> This progestin diol has been prepared by *cis*-hydroxylation of 16-dehydroprogesterone (1), but the reported yields were low.<sup>2</sup> Our initial attempts at selective oxidation of the sterically encumbered  $\text{C}_{16}$ -olefin in 1 with potassium permanganate in aqueous acetone or catalytic osmium tetroxide with excess *N*-methylmorpholine-*N*-oxide gave the desired diol 2, but in low yield (15%-25%). In an attempt to improve the efficiency of this reaction,<sup>3</sup> we used the permanganate phase transfer catalyst, cetyltrimethylammonium permanganate, as an oxidant in dichloromethane.<sup>4</sup> The product isolated after a non-aqueous work up procedure, however, was not the desired  $16\alpha,17\alpha$ -progestin diol 2, but the novel 5-ring D-homosteroid 3.



Information obtained from  $^1\text{H}$  and  $^{13}\text{C}$  NMR, IR and mass spectrometry was useful in the initial assignment of the structure of the novel D-homosteroid.<sup>5</sup> The  $^1\text{H}$  NMR spectrum revealed only two  $\text{CH}_3$  groups; the absence of a singlet downfield of  $\delta$  2.0 suggested that the  $\text{C}_{21}$  methyl group had been lost. There was very little change in the peaks corresponding to the A, B, and C rings of the progestin backbone. The  $^{13}\text{C}$  NMR spectrum contained 21 unique carbon signals, which included two carbonyl carbons and two olefinic carbons. One of the carbonyl  $^{13}\text{C}$  resonances was shifted upfield to  $\delta$  178.2, which suggested that this carbonyl had been transformed from a ketone to an ester. EI mass spectrometry showed a molecular ion at 344, which indicated that the compound had undergone additional oxidation compared to the desired progestin

diol 2. HRMS indicated a molecular formula of  $C_{21}H_{28}O_4$ . The infrared spectrum contained bands for a hydroxyl ( $3559\text{ cm}^{-1}$ ) and two carbonyl functionalities ( $1774$  and  $1664\text{ cm}^{-1}$ ). A crystal suitable for X-ray crystallography was grown by slow evaporation of a  $CH_2Cl_2$  solution of 3 at room temperature. The crystal structure<sup>6</sup> (Figure 1) identified 3 as a novel D-homosteroid with an additional lactone ring bridging the ring-expanded D-ring on the  $\alpha$ -face between  $C_{16}$  and  $C_{17a}$ .

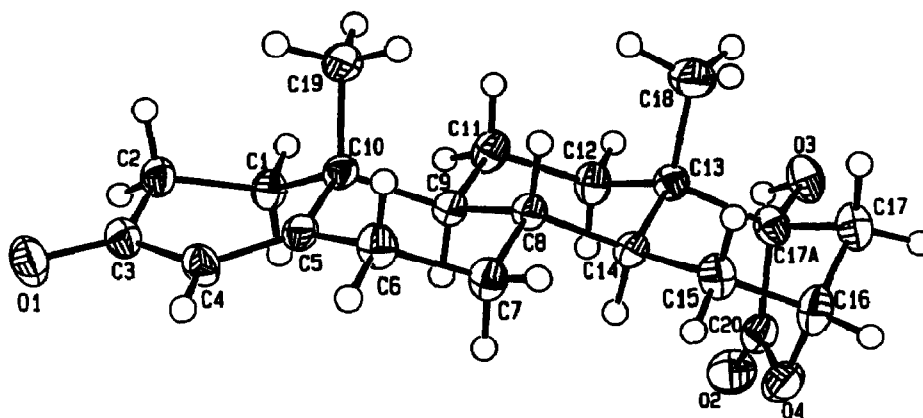


Figure 1. X-ray crystallographic structure of the D-homosteroid 3.

In an attempt to optimize the yield of the D-homo steroid 3, several modifications of the reaction conditions were investigated. We found that forcing reaction conditions (excess CTAP, longer reaction times, or heat) resulted in lower yields of the desired product, presumably due to further oxidative degradation of the progestin backbone. Oxidation using CTAP in a mixture of acetone and 1.0 M potassium hydroxide (1:1 v:v) resulted in formation of diol 2 in 51% yield.

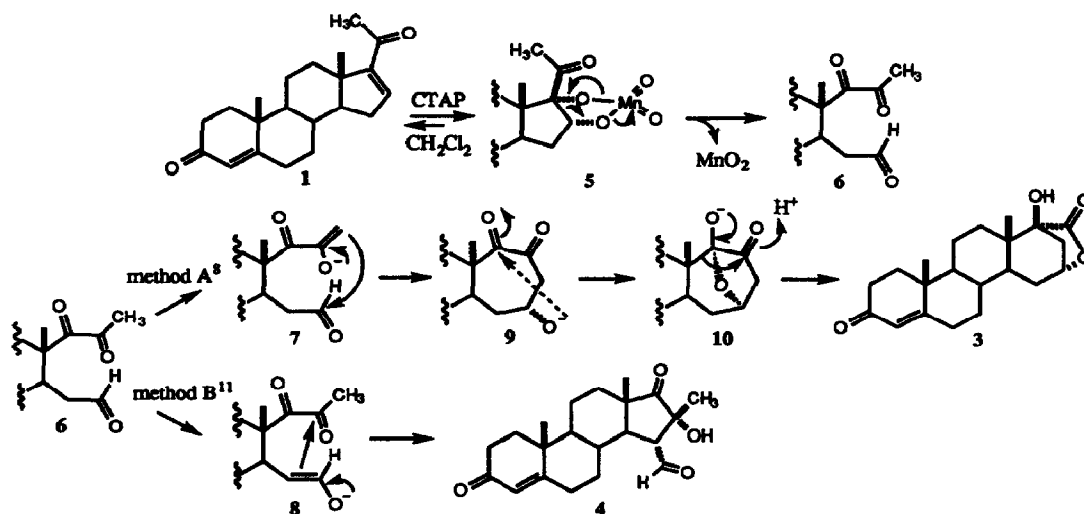
Of particular interest, we found that slow addition of a dichloromethane solution of CTAP via a syringe pump over 8 h to a  $CH_2Cl_2$  solution of 16-dehydroprogesterone resulted in formation of a slightly less polar derivative as the major product (28% based on consumed starting material), together with the D-homo steroid 3 in trace amounts (< 2%). We have identified this less polar derivative as the novel androstane carboxaldehyde 4.<sup>7</sup> The distinguishing features from  $^1H$  NMR spectrum are the aldehydic proton resonance ( $\delta$  9.69), and three large singlets ( $\delta$  1.03, 1.21, 1.45) that can be attributed to methyl groups. A decoupling experiment established the link between the aldehydic proton and the proton on the  $C_{15}$  methine. The  $^{13}C$  NMR shows three resonances ( $\delta$  217.47, 203.94, 199.16) assigned to ketone or aldehydic carbons, and the two olefinic carbons in the A-ring ( $\delta$  169.20, 124.39). The infrared spectrum revealed bands for the hydroxyl ( $3572\text{ cm}^{-1}$ ) and three carbonyl ( $1748$ ,  $1719$ ,  $1665\text{ cm}^{-1}$ ) functionalities. EI mass spectrometry identified the molecular ion as 344, placing this compound in the same oxidation state as the D-homosteroid 3. HRMS indicated the molecular formula to be  $C_{21}H_{28}O_4$ . The structure assigned to compound 4 is consistent with all of these data. This novel compound is especially interesting, as it provides a handle for further manipulation at  $C_{15}$  in the androstane backbone, a site that is difficult to access through conventional methods.

The structure assigned to 4 contains two new stereogenic centers at  $C_{15}$  and  $C_{16}$ . The stereochemistry at these centers was defined via nuclear Overhauser effect (NOE)  $^1H$  NMR studies. Irradiation of the  $C_{15}$

methine proton produced an NOE enhancement of 4.6% at the C<sub>20</sub> methyl group, 5.0% at the C<sub>18</sub> methyl group, and 3.3% of the aldehydic resonance. Irradiation of the C<sub>20</sub> methyl group produced a large NOE enhancement of 5.9% at the C<sub>15</sub> methine proton and a small enhancement of 1.4% at the C<sub>18</sub> proton. This establishes that the C<sub>15</sub> methine proton, the C<sub>18</sub> methyl group and the C<sub>20</sub> methyl group are on the same side of the molecule. Since the C<sub>13</sub> stereogenic center (site of C<sub>18</sub> methyl attachment) has not changed in the transformation from 1 to 4, we assign these groups on the β-face of the molecule. This assignment is also consistent with the large coupling constant (12.4 Hz) observed between the anti-disposed C<sub>15</sub> and C<sub>14</sub> methine protons.

It is intriguing to speculate on a possible mechanism for the formation of the novel steroids 3 and 4. In our proposed mechanism (Scheme I), under non-aqueous conditions the breakdown of the cyclic manganese (VI) ester 5 results in scission of the C<sub>16</sub>-C<sub>17</sub> bond and opening of the D-ring. The resulting diketone 6 is able to enolize at two different sites. Under reaction conditions in which the permanganate concentration is high (method A<sup>8</sup>), the C<sub>21</sub> enolate of the diketone (7) attacks the aldehyde to form the seven-membered ring intermediate 9; subsequent attack of the alkoxide at the C<sub>17</sub> carbonyl forms the 5-membered ketolactol 10; finally, ring contraction of 10 is accomplished through a mechanism analogous to the benzylic acid rearrangement<sup>9</sup> forming the D-homosteroid 3. The ring contraction of the D-ring resembles a reported ring contraction of the C-ring in an androstane 11,12-diketone by a benzylic acid rearrangement under basic conditions.<sup>10</sup> If the concentration of permanganate ion is low (method B<sup>11</sup>), the C<sub>15</sub>-enolate 8 attacks at the C<sub>20</sub> carbonyl, affording the novel androstane 4. To investigate whether androstane 4 could be converted to the D-homosteroid 3 under basic conditions we treated it with 1,8-diazabicyclo[5.4.0]undec-7-ene in CH<sub>2</sub>Cl<sub>2</sub>. This afforded a 4:1 mixture of 4 with its C<sub>16</sub> epimer, but formation of D-homosteroid 3 was not observed.

Scheme I



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3. We found that stoichiometric osmium tetroxide in pyridine converts 16-dehydroprogesterone to progestin diol **2** in 82% yield with high regio and stereoselectivity. A detailed experimental section is given in ref. 1.
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5. Spectroscopic data for **3**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.05 (s, 3 H, 19- $\text{CH}_3$ ), 1.19 (s, 3 H, 18- $\text{CH}_3$ ), 2.59 (s, 1 H, OH), 4.81 (dd,  $J = 5.16, 5.24$  Hz, 1 H, 16 $\beta$ -CH), 5.72 (s, 1 H, 4-CH);  $^{13}\text{C}$  NMR (125 MHz)  $\delta$  199.58, 178.25, 170.47, 123.85, 79.87, 75.28, 53.17, 44.26, 39.98, 39.21, 38.75, 35.35, 34.11, 33.91, 32.54, 30.76, 29.44, 27.97, 20.08, 17.46, 12.17; IR ( $\text{CHCl}_3$ ) 3559, 1774, 1664  $\text{cm}^{-1}$ ; MS (EI, 70 eV)  $m/z$  344 ( $\text{M}^+$ ), 329, 316, 273. HRMS Calcd for  $\text{C}_{21}\text{H}_{28}\text{O}_4$  344.1988, found 344.1985. Anal. Calcd for  $\text{C}_{21}\text{H}_{28}\text{O}_4$ : C, 73.23; H, 8.19. Found: C, 73.02; H, 8.23.
6. Compound **3** crystallizes in an orthorhombic system of space group  $\text{P}2_12_12_1$  with 4 molecules in a unit cell of dimensions  $a = 6.477$  (2),  $b = 12.742$  (5),  $c = 21.121$  (8),  $V = 1743$  (1)  $\text{\AA}^3$ . Using a total of 1993 reflections measured on an Enraf-Nonius diffractometer using monochromated  $\text{MoK}\alpha$  radiation, the structure was solved by a direct method (SHELXS-86), and a differential Fourier method (for H), and refined by least squares on F2 using SHELXL-93. The final R factor was 4.5%. Final crystallographic coordinates have been deposited in the Cambridge Crystallographic Data Center.
7. Spectroscopic data for **4**:  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  1.03 (s, 3 H, 19- $\text{CH}_3$ ), 1.21 (s, 3 H, 18- $\text{CH}_3$ ), 1.45 (s, 3 H, 20- $\text{CH}_3$ ), 2.60 (dd,  $J = 12.4, 4.2$  Hz, 1 H, 15-CH), 5.74 (s, 1 H, 4-CH), 9.69 (d,  $J = 4.2$  Hz, 1 H, aldehyde);  $^{13}\text{C}$  NMR (125 MHz)  $\delta$  217.47, 203.94, 199.16, 169.20, 124.39, 79.64, 58.24, 53.77, 48.18, 48.10, 38.73, 35.79, 34.99, 33.97, 32.40, 31.95, 31.83, 25.80, 20.04, 17.55, 15.40; IR ( $\text{CHCl}_3$ ) 3572, 1748, 1719, 1665  $\text{cm}^{-1}$ ; MS (EI, 70 eV)  $m/z$  344 ( $\text{M}^+$ ), 326, 286, 273, 255, 230, 107. HRMS Calcd for  $\text{C}_{21}\text{H}_{28}\text{O}_4$  344.1988, found 344.1985.
8. Procedure for method A: A  $\text{CH}_2\text{Cl}_2$  solution of CTAP (0.310 g, 0.77 mmol) was added over 10 min to a stirred solution of 16-dehydroprogesterone (0.200 g, 0.64 mmol) in 10 mL of dry  $\text{CH}_2\text{Cl}_2$  and the resulting brown solution stirred for 2 h at RT. Evaporation of the  $\text{CH}_2\text{Cl}_2$  followed by addition of 100 mL ether resulted in a heterogeneous mixture of clear liquid and brown solid. The mixture was filtered through a plug of celite and  $\text{MgSO}_4$  (10 g of each) and the plug was washed twice with 50 mL portions of ether. The ether layer was evaporated to provide 110 mg of white solid. Flash chromatography (1:1 hexanes/EtOAc) afforded 0.062 g of **3** as a white solid (mp 236-238  $^\circ\text{C}$ ) and 0.038 g of 16-dehydroprogesterone. The yield based on consumed starting material was 35%.
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10. Kurath, P. *J. Org. Chem.* **1967**, *32*, 3626.
11. Procedure for method B: A 30 mL  $\text{CH}_2\text{Cl}_2$  solution of CTAP (0.149 g, 0.37 mmol) was added over 8 h via syringe pump to a stirred solution of 16-dehydroprogesterone (0.096 g, 0.31 mmol) in 100 mL of dry  $\text{CH}_2\text{Cl}_2$  and the resulting brown solution stirred for 2 h at RT. The workup was identical to Method A. Flash chromatography (1:1 hexanes/EtOAc) afforded 0.015 g of **4**, as a white solid (mp 219-221  $^\circ\text{C}$ ), 0.0013 g of **3**, and 0.046 g of 16-dehydroprogesterone. The yield of **4** based on consumed starting material was 28%.

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